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(21) International Application Number: PCT/US98/03638 (22) International Filing Date: 25 February 1998 (25.02.98) (30) Priority Data: 60/039,111 26 February 1997 (26.02.97) US (71)(72) Applicants and Inventors: KHAW, Ban-An [US/US]; 69 Meadowview Road, Milton, MA 02186 (US). NARULA, Jagat [IN/US]; Apartment #1, 116 Longwood Avenue, Brookline, MA 02146 (US). (74) Agents: HEINE, Holliday, C. et al.; Weingarten, Schurgin, Gagnebin & Hayes LLP, Ten Post Office Square, Boston, MA 02109 (US).		(81) Designated States: CA, JP, US, European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). Published <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
(54) Title: SIGNAL ENHANCEMENT OF BISPECIFIC ANTIBODY-POLYMER PROBE FOR IMMUNOASSAY USE (57) Abstract <p>An immunoassay method including reacting a sample from a patient with a bispecific antibody, wherein the bispecific antibody includes one antibody specific for a compound to be detected and a second antibody specific for a compound foreign to said patient sample, and subsequently reacting the patient sample with a polymer probe, wherein the polymer probe includes a compound recognized by the second antibody in the bispecific antibody complex and further includes at least two detectable signals; the bispecific antibody; and the polymer probe of the immunoassay method are disclosed.</p>		

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5 SIGNAL ENHANCEMENT OF BISPECIFIC ANTIBODY-POLYMER PROBE
FOR IMMUNOASSAY USE

CROSS REFERENCE TO RELATED APPLICATIONS

10 STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR
DEVELOPMENT

BACKGROUND OF THE INVENTION

15 An immunoassay utilizes antibodies to detect a compound
of choice. However, the sensitivity of this detection is
generally limited by the amount of signal that can be carried
either on the antibody, for a direct binding assay, or on the
probe compound, in a competitive inhibition assay. For
20 example, in existing immunoassays, such as
radioimmunoassay, ELISA, immunofluorescent assays or
immunochemiluminescent assays, too many signal entities, such
as radioisotopes, horse radish peroxidase or alkaline
phosphatase, attached to the detection moieties invariably
25 inactivate the antibody or denature the antigen and change
the property of the detection probe. Therefore, in order
to obtain more signal, additional antibody or probe must be
added. This, in turn, reduces the sensitivity of the assay,
the capability of the assay to detect minute quantities of
30 the compound in question.

For all existing immunoassays, there is lag time for the
compound of interest to reach a high enough concentration in
the serum to become detectable for diagnostic purposes. In
the case of heart attacks, there is a delay of 4-6 hours from
35 the onset of chest pain until the diagnostic detection of CK-
MB, Troponin-T or I is possible. Myoglobin is detectable
earlier, but its specificity is low. If there were an assay
that could detect very minute increases of these indicator
compounds in the blood at an earlier point in time, then

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therapeutic intervention could be started earlier and thereby bring about greater myocardial salvage. In the case of cancer detection, where, e.g., tumor associated antigens related to breast cancer or colon cancer, etc., are detected, treatment might be more effective if minute elevations of these antigens could be detected at an early stage. Therefore, there is a need to increase the sensitivity of the assay without adversely affecting the specificity of the assay system.

SUMMARY OF THE INVENTION

The invention is directed to a method to increase the sensitivity of an immunoassay, by at least 10,000 fold, without losing specificity. This improvement is achieved by the use of a bispecific antibody complex and a unique detection signal probe capable of recognizing the bispecific antibody complex.

In one aspect, the invention features an immunoassay method including reacting a sample from a patient with a bispecific antibody, wherein the bispecific antibody includes one antibody specific for a compound to be detected and a second antibody specific for a compound foreign to said patient sample, and subsequently reacting the patient sample with a polymer probe, wherein the polymer probe includes a compound recognized by the second antibody in the bispecific antibody complex and further includes at least two detectable signals. The invention also features the bispecific antibody and the polymer probe of the method of the invention. Preferably, the sample from the patient is a blood or serum sample; the bispecific antibody includes an antimyosin antibody and an antibody against DTPA; and the polymer probe is a polylysine polymer and includes DTPA and at least six HRP as the detectable signal compounds.

BRIEF DESCRIPTION OF THE DRAWINGS

Other features and advantages of the invention will be apparent from the following description of the preferred

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embodiments thereof and from the claims, taken in conjunction with the accompanying drawings, in which:

Fig. 1a shows a standard ELISA according to the prior art;

5 Fig. 1b shows an immunoassay according to the invention; and

Fig. 2 is a graph showing competitive inhibition curves using standard ELISA (R11D10), bispecific antibody complex with standard secondary antibody for signal production (BiMab (Ab-HRP)), and the method according to the invention (BiMab(PL-DTPA-HRP)).

DETAILED DESCRIPTION OF THE INVENTION

15 The invention is directed to the development of a new approach to the use of bispecific antibodies in immunoassays. The new specific antibody comprises one antibody specific for the compound associated with the pathological state to be detected and another antibody to a chemical or reporter compound that is not found naturally in man. These two are

20 chemically or genetically linked. The bispecific antibody complex constitutes the first line of interaction with the compound one is attempting to detect. Normally many antibodies must react with the compound to enable development of sufficient signal intensity for detection. However, in

25 the method of the invention, a novel detection probe is used, made up of any type polymer, such as polylysine or other polyamino acid, that is amenable to attachment of signal reagents and reporter compounds. The amount of signal reagent that can be used in a given assay is limited only by

30 the size of the polymer. Only a few molecules of the detection probe are therefore needed to provide this signal. The signal probe is extremely versatile as any type of signal producing compound such as radioactivity, chemical color producing enzymes or fluorescent probes can be attached to

35 the polymer backbone. Signal amplification is not limited by the nature of the bispecific antibody complex itself.

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Therefore, the immunoassay sensitivity can be amplified by at least 10,000-fold compared to conventional immunoassays or immunosandwich assays. Since early detection of many pathological states, such as acute myocardial infarction and cancer, is limited by the sensitivity of immunoassays to detect minute elevations of the pathologically associated compounds, a method and compounds of the invention will enable diagnosis of disease states at a much earlier time than previous assays, which may allow for better therapeutic intervention.

Another advantage of the method of the invention is the versatility for adaptation to any antibody. For example, the method could be adapted to detect troponin-I or T by using the antibody specific for troponin-I or T attached to a second antibody, such as the antibodies shown herein, that recognizes the detector probe. If higher sensitivity is necessary, the polymer probe could be generated to carry higher numbers of signal compounds. Furthermore, the polymer probe can include any kind of signal compound, such as radioisotope, fluorescent, or paramagnetic linked signal compounds.

All previously existing ELISA radioimmunoassays, dipstick assays for cancer, pregnancy, serum enzymes and probes and any assays utilizing antibodies could be modified according to the method of the invention to provide enhanced sensitivity. In addition, *in vivo* application to enhance target signal by using the method of the invention is also possible.

The following examples are presented to illustrate the advantages of the present invention and to assist one of ordinary skill in making and using the same. These examples are not intended in any way otherwise to limit the scope of the disclosure.

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EXAMPLE I

Serum immunoassays for intracardiac contractile proteins constitute the mainstay for detection of myocyte necrosis associated with various cardio-vascular disorders. However, myosin heavy chain (MHC) fragments can be detected by immunoassay only after 48 h from the onset of chest pain. To enhance immunodetection of MHC, monoclonal antibody (MAb) R11D10 specific for cardiac MHC was covalently linked to MAb 4G4-1D5 specific for DTPA. The probe consisted of DTPA-modified polylysine (28:1 molar ratio) covalently linked to horse-radish peroxidase (6 moles/mole polylysine) (PL-DTPA-HRP). Porcine cardiac myosin (PCM, 1 μ g/ml) was used to coat the microtiter wells. After overnight incubation and washing, three times, 50 μ l each of 5 μ g/ml BiMAb or MAb and serial dilutions of PCM (0.001 to 100 μ g/ml) or 50 μ l of serial dilutions (1/1 to 1/10000) of patient sera pre-incubated for 1 h at 37°C were added and incubated for 2 h at 37°C. After washing, the wells were incubated with goat-antimouse IgG-HRP or PL-DTPA-HRP for 2 h. A chromogen, dinitrobenzidine was used to develop the assay. The affinity of BiMAb and R11D10 were the same at 1.5 x 10⁹L/mole. The sensitivity of BiMAb was 0.5 ng, whereas that of R11D10 was 0.5 μ g (1 μ g/ml). BiMAb developed with the conventional goat anti-mouse IgG-HRP had a sensitivity of 0.05 μ g. Therefore, BiMAb assay has a 1000 fold increase in sensitivity compared to the conventional immunoassay in the sera of 3 heart transplant patients. Using the BiMAb assay, 2.5, 1.25 and 1.3 ng MHC/50 μ l serum at 1/10³ dilution, were detected. This BiMAb technology can be used in RIA or ELISA by interchanging the HRP probe for radiolabeled probe and should provide more specific in vitro diagnosis of acute myocardial infarction since detection of MHC is not feasible at the present time of day 1 of myocardial infarction by conventional immunoassays.

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EXAMPLE II

In a subsequent experiment the DTPA-modified polylysine probe of Example I was covalently linked to 12 moles of horse-radish peroxidase per mole of polylysine. The results of the study show that the sensitivity of the bispecific assay of the invention (10^{-5} to $100\mu\text{g/ml}$) was at least 10,000 fold better than the conventional immunoassay ($0.1\mu\text{g/ml}$).

While the present invention has been described in conjunction with a preferred embodiment, one of ordinary skill, after reading the foregoing specification, will be able to effect various changes, substitutions of equivalents, and other alterations to the compositions and methods set forth herein. It is therefore intended that the protection granted by Letters Patent hereon be limited only by the definitions contained in the appended claims and equivalents thereof.

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CLAIMS

What is claimed is:

1. An immunoassay method comprising

5 reacting a sample from a patient with a bispecific antibody, said bispecific antibody comprising one antibody specific for a compound to be detected and

10 a second antibody specific for a compound foreign to said patient sample; and

subsequently reacting said sample with a polymer probe, said polymer probe comprising

15 a polymer backbone, attached to said polymer backbone, a compound recognizable by said second antibody in said bispecific antibody, and

at least two detectable signal compounds further attached to said polymer backbone.

20 2. The immunoassay method of claim 1, wherein said polymer probe comprises at least ten detectable compounds.

25 3. The immunoassay method of claim 1, wherein said detectable signal in said polymer probe is selected from the group consisting of radioisotope, fluorescent probe and paramagnetic probe.

4. The immunoassay method of claim 1, wherein said sample from said patient is a blood or serum sample; said bispecific

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antibody comprises an antimyosin antibody and an antibody against DTPA; and said polymer probe is a polylysine polymer and comprises DTPA and at least 6 HRP as said detectable signal compounds.

5

5. A bispecific antibody for use in an immunoassay comprising

one antibody specific for a compound to be detected in said immunoassay; and

10

a second antibody specific for a compound foreign to a sample to be assayed in said immunoassay.

6. A polymer probe comprising

a polymer backbone;

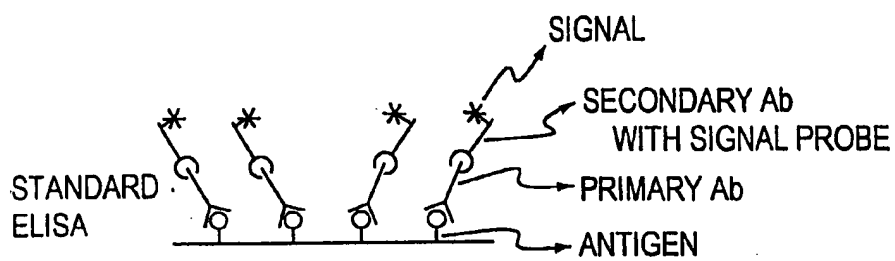
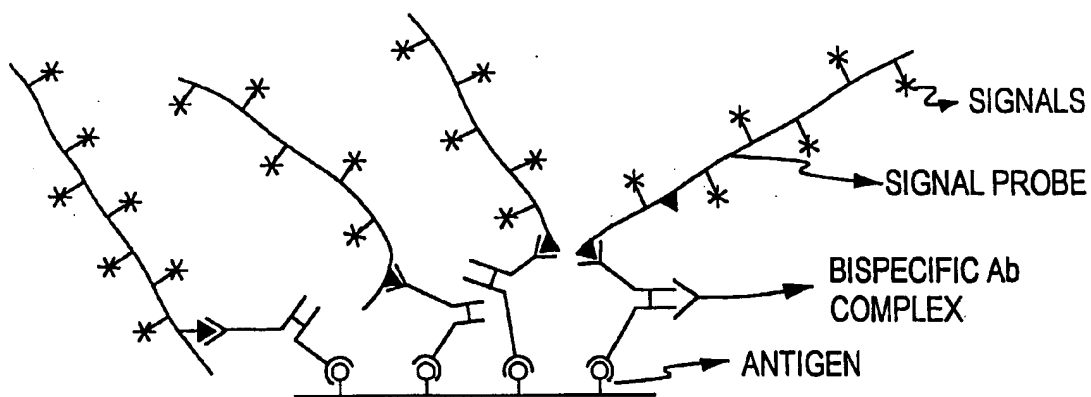
15

a compound recognizable by an antibody in a bispecific antibody attached to said polymer backbone, and at least two detectable signal compounds attached to said polymer backbone.

20

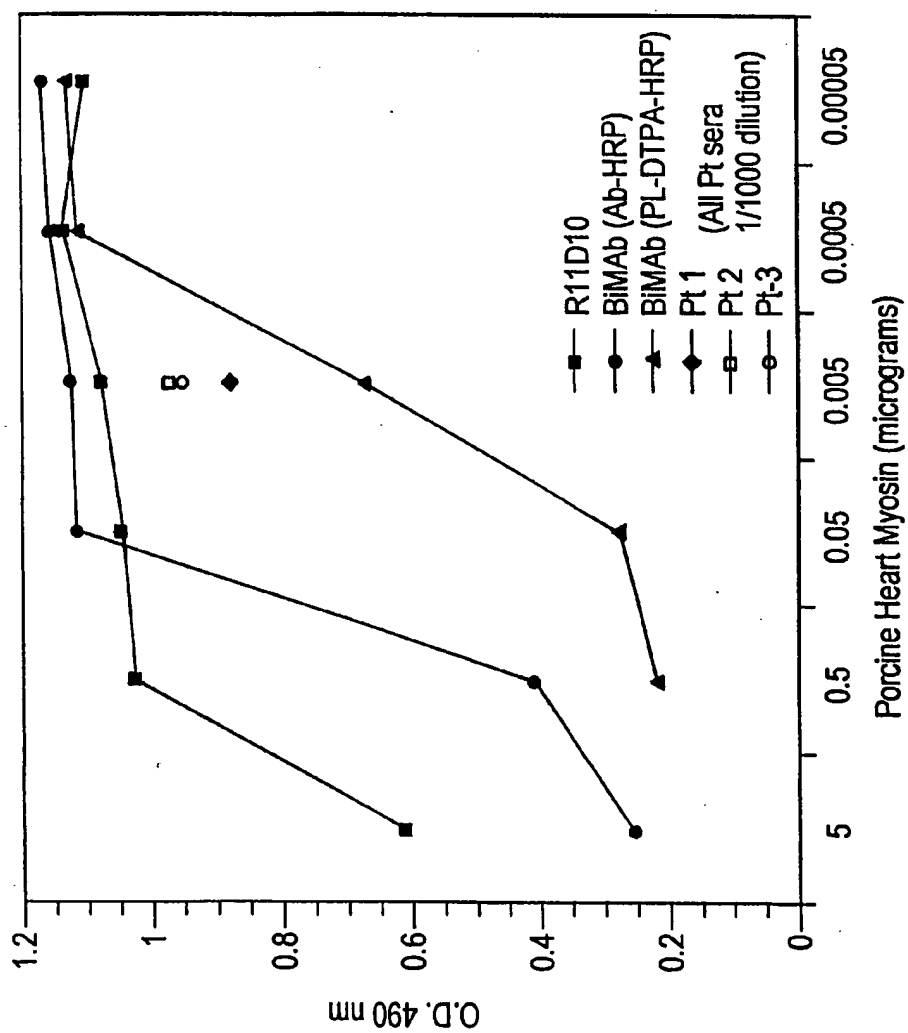
a compound recognizable by said second antibody in said bispecific antibody and at least two detectable signal compounds.

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**FIG. 1a****FIG. 1b**

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**FIG. 2**

SUBSTITUTE SHEET (RULE 26)

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US98/03638

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : Please See Extra Sheet.

US CL : 436/535, 540, 542; 435/7.92; 530/387.3, 817; 930/290

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 436/535, 540, 542; 435/7.92; 530/387.3, 817; 930/290

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, MEDLINE, BIOSIS CAPLUS EMBASE JAPIO PATOSWO, PATOSEP

diethylenetriamine pentaacetic acid, bispecific antibody, polymer backbone, antibody, polylysine

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	TORCHILIN, V.P., et al. The antibody linked chelating polymers for nuclear therapy and diagnostics. Crit. Rev. Therap. Drug Carrier Syst. 1991. Vol. 7, No. 4, pages 275-308, see entire document.	1-6
Y	WO 94/12196 A1 (TANOX BIOSYSTEMS, INC.) 09 June 1994, see entire document.	1-6
Y	US 5,482,698 A (GRIFFITHS et al.) 09 January 1996, see entire document.	1-6
Y, P	US 5,698,178 A (GOLDENBERG et al.) 16 December 1997, see entire document.	1-6
Y	US 5,332,567 A (GOLDENBERG et al.) 26 July 1994, see entire document.	1-6

☒ Further documents are listed in the continuation of Box C.
 ☐ See patent family annex.

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International application No.
PCT/US98/03638

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US 5,591,828 A (BOSSLET et al.) 07 January 1997, see entire document.	1-6
Y	ROSEBROUGH, S.F. Two-step immunological approaches for imaging and therapy. Q. J. Nucl. Med. September 1996. Vol. 40 pages 234-251, see entire document.	1-6
Y,P	VUILLEZ, J., et al. Two-step immunoscintigraphy for non-small-cell lung cancer staging using a bispecific anti-CEA/anti-indium-DTPA antibody and an indium-111-labeled DTPA dimer. J. Nuc. Med. 1997. Vol. 38, No. 4, pages 507-511, see entire document.	1-6
Y	KRANENBORG, M.H.G.C., et al. Development and characterization of anti-renal cell carcinoma X antichelate bispecific monoclonal antibodies for two-phase targeting of renal cell carcinoma. Canc. Res. 01 December 1995. Vol. 55, 23 Supplement, pages 5864s-5867s, see entire document.	1-6
Y	DEVYS, A., et al. Comparative targeting of human colon-carcinoma multicell spheroids using one- and two-step (bispecific antibody) techniques. Int. J. Cancer. 1996. Vol. 67, pages 883-891, see entire document.	1-6

Form PCT/ISA/210 (continuation of second sheet)(July 1992)*

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A. CLASSIFICATION OF SUBJECT MATTER:

IPC (6):

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